LETTER TO THE EDITOR

Novel gene therapy for rheumatoid arthritis with single local injection: adeno-associated virus-mediated delivery of A20/TNFAIP3

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Dear Editor,

Multiple experiments have established TNF alphainduced protein 3 (A20/TNFAIP3) as a critical regulator associated with rheumatoid arthritis (RA) [1, 2]. The lack of TNF- α -induced protein 3 (A20) promotes the NOD-like receptor protein 3 (NLRP3) inflammasome and induces spontaneous arthritis, while increase of A20 reduces the secretion of IL-1 β and favors immunological tolerance [3, 4]. Hence, we investigate the feasibility of recombinant adeno-associated virus 6 (rAAV6)-mediated A20 gene therapy in a collagen-induced arthritis (CIA) model.

We first investigated cytomegalovirus (CMV) promoter-regulated gene delivery (Additional file 1: Fig. S1). rAAV6 (Additional file 1: Fig. S1a, b) was transfected into 293 T cells (Additional file 1: Fig. S2a) to enhance A20 expression (Additional file 1: Fig. S2b). A 10 µl of volume rAAV6-CMV-A20 containing 1×10^{12} viral genomes (vg) was injected into the left knee, ankle, and tarsal area of CIA mice, while the same dose of rAAV6-CMV-EGFP was injected into the right side (Additional file 1: Fig. S2c). rAAV6 was widely distributed in various cell types (Additional file 1: Fig. S2d) and significantly enhanced A20 expression until 5 weeks after injection (Fig. 1a). Notably, rAAV6-CMV-A20 decreased the clinical arthritis score, paw thickness and total porosity (P<0.001), increased the bone volume-to-tissue volume ratio (BV/ TV, P<0.001), trabecular number (Tb.N, P=0.008), and trabecular thickness (Tb.Th, P=0.002) (Fig. 1b–d, Additional file 1: Fig. S2e–j), and suppressed pannus formation, bone erosion, and cartilage destruction (Fig. 1e). We also found that rAAV6-CMV-A20 significantly suppressed the expression of NLRP3, caspase-1, and IL-1 β (Additional file 1: Fig. S2k–m). Thus, the results verified our hypothesis that rAAV6-mediated A20 overexpression is an effective RA therapy.

The CMV promoter may attract safety concerns. Therefore, considering that macrophages and fibroblastlike synoviocytes account for the primary components of synovial tissues, especially in RA, we tested which cell type predominantly overexpressed A20. We constructed rAAV6-COL1a-A20 (Additional file 1: Fig. S1c) targeting fibroblast-like synoviocytes and rAAV6-SP146-A20 (Additional file 1: Fig. S1e) targeting macrophages [5]. A 10 μl volume of rAAV6-COL1α-A20 or rAAV6-SP146-A20 $(1 \times 10^{12} \text{ vg})$ was injected into the left knee, ankle, and tarsal area, while the same dose of rAAV6-COL1 α -EGFP (Additional file 1: Fig. S1d) or rAAV6-SP146-EGFP (Additional file 1: Fig. S1f) was injected into the same areas on the right side (Additional file 1: Fig. S3a). In contrast to rAAV6-COL1α-EGFP, rAAV6-COL1α-A20 exhibited nearly no effect on suppressing RA symptoms. However, rAAV6-SP146-A20 showed anti-rheumatic



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with rAAV6-CMV-A20 or rAAV6-SP146-A20. I Immunofluorescence staining of A20 and F4/80 in joint specimens injected with rAAV6-SP146-A20. CMV cytomegalovirus, NLRP3 NOD-like receptor protein 3

therapeutic effects compared with rAAV6-SP146-EGFP. Similar to rAAV6-CMV-A20, rAAV6-SP146-A20 reduced the clinical arthritis score, paw thickness (Fig. 1f, g; Additional file 1: Fig. S3b, c), total porosity (Additional file 1: Fig. S3d), bone erosion activity (Additional file 1: Fig. S3e–g), pannus formation, and cartilage destruction (Fig. 1h, i). rAAV6-SP146-EGFP was well transfected into RAW264.7 cells (Fig. 1j) and exhibited dominant distribution in macrophages (Additional file 1: Fig. S3h). rAAV6-SP146-A20 significantly enhanced A20 expression (Fig. 1k, l) and the collective results further indicated the therapeutic benefits of rAAV6-mediated A20 overexpression. Importantly, macrophages were found to be responsible for the rAAV6-mediated A20 overexpression.

The anti-rheumatic benefits of rAAV6-SP146-A20 were further verified by simultaneous injection of rAAV. CIA mice were injected with rAAV6-SP146-EGFP or rAAV6-SP146-A20 on both sides $(1 \times 10^{12} \text{ vg}, \text{Additional})$ file 1: Fig. S3i). As expected, rAAV6-SP146-A20 significantly relieved the arthritis symptoms (Additional file 1: Fig. S3j–o, Fig. S4a–c). Furthermore, we tested whether the therapeutic effect was dependent on viral genomes. CIA mice were injected with rAAV6-SP146-EGFP at

a dose of 1×10^{12} vg (EGFP) or rAAV6-SP146-A20 at a dose of 1×10^8 vg (E8), 1×10^{10} vg (E10), or 1×10^{12} vg (E12, Additional file 1: Fig. S3p). The results revealed that the protective effect of rAAV6-SP146-A20 was dose-dependent. The E12 group exhibited the lowest clinical arthritis score and increases in paw thickness, bone erosion activity, pannus formation, and cartilage destruction (Additional file 1: Figs. S3q–v, S4d–e). As the treatment dose increased, so did the expression of A20, while NLRP3, caspase-1, and IL-1 β were gradually inhibited (Additional file 1: Figs. S4f–g).

Our hypothesis that A20 overexpression is a reasonable strategy for the treatment of RA was initially evidenced. We not only demonstrated the therapeutic effects against RA of ubiquitous CMV promoter-driven delivery of A20, but more importantly, we identified the key role of macrophage-like synovial cells in responding to rAAV6mediated gene delivery of A20.

Abbreviations

A20/TNFAIP3: TNF-α-induced protein 3; BV/TV: Bone volume-to-tissue volume ratio; CMV: Cytomegalovirus; CIA: Collagen-induced arthritis; NLRP3: NOD-like receptor protein 3; RA: Rheumatoid arthritis; Tb.N: Trabecular number; Tb.Th: Trabecular thickness.

Supplementary Information

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Additional file 1. Materials and methods. Fig. S1: Construction of rAAV expression plasmids. Fig. S2: Therapeutic effect of rAAV-CMV-A20. Fig. S3: Verification of the therapeutic effect of rAAV6-SP146-A20. Fig. S4: rAAV6-SP146-A20 inhibits NLRP3-mediated inflammation in CIA mice.

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Not applicable.

Author contributions

QZ performed the experiments, analyzed the data, and wrote the manuscript. FXY established rAAV6. YLW, CYY, NCL, and XZ assisted with the cell experiments and data analysis. PMZ and ZYW designed rAAV6 and supervised the development of rAAV6. JL conceived and designed the study. All authors read and approved the final version of the manuscript.

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Availability of data and materials

All data needed to evaluate the conclusions in the paper are present in the paper or the Additional files. The data are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

Fang-Xing Yu, Pi-Ming Zhao, and Zhong-Ya Wang (CureGenetics Co., Ltd.) are responsible for the encapsulation and identification of the rAAV required for the experiment. The authors declare that they have no competing interest for this work.

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